

Pathogenesis of deep endometriosis

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The pathophysiology of (deep) endometriosis is still unclear. As originally suggested by Cullen, change the definition “deeper than 5 mm” to “adenomyosis externa.” With the discovery of the old European literature on uterine bleeding in 5%–10% of the neonates and histologic evidence that the bleeding represents decidual shedding, it is postulated/hypothesized that endometrial stem/progenitor cells, implanted in the pelvic cavity after birth, may be at the origin of adolescent and even the occasionally premenarcheal pelvic endometriosis. Endometriosis in the adolescent is characterized by angiogenic and hemorrhagic peritoneal and ovarian lesions. The development of deep endometriosis at a later age suggests that deep infiltrating endometriosis is a delayed stage of endometriosis. Another hypothesis is that the endometriotic cell has undergone genetic or epigenetic changes and those specific changes determine the development into deep endometriosis. This is compatible with the hereditary aspects, and with the clonality of deep and cystic ovarian endometriosis. It explains the predisposition and an eventual causal effect by dioxin or radiation. Specific genetic/epigenetic changes could explain the various expressions and thus typical, cystic, and deep endometriosis become three different diseases. Subtle lesions are not a disease until epi(genetic) changes occur. A classification should reflect that deep endometriosis is a specific disease. In conclusion the pathophysiology of deep endometriosis remains debated and the mechanisms of disease progression, as well as the role of genetics and epigenetics in the process, still needs to be unraveled. [Fertil Steril® 2017;108:872–85. ©2017 by American Society for Reproductive Medicine.]

Key Words: Deep endometriosis, pathogenesis, classification, heredity, genetics, epigenetics, neonatal menstruation

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At the turn of the 19th century Cullen (1–3) described 10 different sites in the pelvis where he found the presence of “uterine mucosa.” When located in the rectovaginal septum he called it a rectovaginal adenomyoma. Meyer (4) and Gruenwald (5) suggested that this was caused by metaplasia. Later Sampson (6, 7) suggested retrograde menstruation with the tubal transport of endometrial cells as etiology. When retrograde menstruations was found in almost all women (8, 9) speculation started why not all women developed endometriosis. It remains debated whether these lesions should be considered either as initial lesions after implantation or as “a physiologic” phenomenon occurring intermittently in all women (10).

Deep endometriosis was described in the early nineties as adenomyosis externa (11) with endometrial glands and stroma in fibromuscular tissue. Because the glandular activity was more in phase ($\leq 75\%$) with the menstrual cycle at depths >5 mm deep endometriosis was defined as lesions >5 mm in the peritoneum (12). This definition seemed consistent with the concept that deep endometriosis had escaped from the high steroid concentrations in peritoneal fluid (PF) (13) (Supplemental Fig. 1). At present we realize that this definition was a mistake and should be abandoned. The 5-mm definition permits the inclusion of slightly deeper typical lesions. It would have been preferable to define deep endometriosis as adenomyosis externa. With this latter definition most deep endometriosis le-

sions are unique (occasionally 2 and rarely 3) and big (mostly >1 cm in diameter). These deep endometriosis lesions seem to develop as a benign tumor, preferentially in the pouch of Douglas, with extension toward the uterine artery or the ureters, with a preferential invasion into the muscle of the bowel wall or the diaphragm, but not into the fat. These adenomyosis externa lesions occasionally invade nerves (14) and have some neurotropic effect (15, 16). In most bowel lesions lymph nodes are invaded (17, 18).

To avoid confusion metaplasia and genetic or epigenetic changes are defined as follows. Metaplasia (19) is the reversible transformation of one differentiated cell type to another differentiated cell type. This may be part of a normal maturation process or caused by some abnormal stimulus. If the stimulus causing metaplasia is removed, tissues return to their normal pattern of differentiation. Genetic and epigenetic changes are permanent heritable changes in DNA sequence or in gene function not associated with changes in DNA sequence (20).

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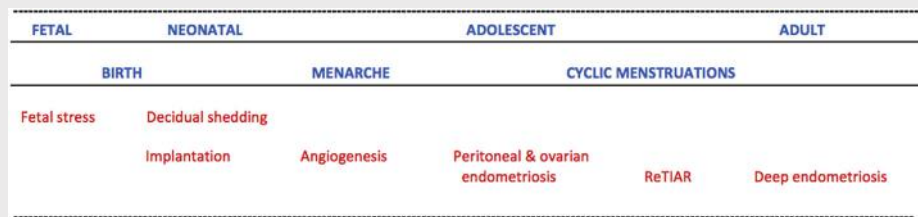
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FIGURE 1



Life cycle of early onset endometriosis. ReTIAR = “recurrent tissue injury and repair” leading to adenomyotic and fibrotic changes of deep endometriosis.

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Deep endometriosis as the only form of disease in absence of other endometriotic lesions was present in only 6.5% (21). The high correlation between the presence of deep endometriosis and the presence of peritoneal and ovarian endometriosis raises the question whether they are three different entities with a common or different pathogenesis. That peritoneal, ovarian, and rectovaginal endometriotic lesions represent three clinically separate disease entities with a different pathogenesis had already been suggested 20 years ago (22). Are ovarian endometrioma (OMA) and rectovaginal endometriosis phenotypes of a progressive disease with as main driver recurrent menstrual bleeding causing repeated tissue injury and repair (ReTIAR) or are they caused by (epi)genetic changes? As the pathophysiology of deep endometriosis remains debated and in absence of clear evidence the two visions on pathogenesis were developed separately.

HYPOTHESIS I: PATHOGENESIS OF EARLY ONSET ENDOMETRIOSIS BY NEONATAL UTERINE BLEEDING WITH THE CYCLIC MENSTRUATION AS DRIVING MECHANISM FOR ADENOMYOTIC FORMATION

The Forgotten Menstruation

At birth the endometrial cell in the neonatal uterus expresses a variable response to maternal P. In a classic autopsy study of the neonatal endometrium the Harvard pathologists Ober and Bernstein (23) described that in 5% of the neonates the endometrium is P responsive and responds with decidualization and menstrual changes. In 95% of the neonates, however, the endometrium responds with weak proliferation or secretory changes despite the high maternal P levels during pregnancy. These histologic findings are in agreement with the occurrence of neonatal menstruation during the first week after birth in approximately 5% of the neonates (24) (Fig. 1). The discovery of a large European scientific literature on neonatal menstruation and the recent reports on premenarcheal endometriosis, including OMAs, raised the question whether neonatal uterine bleeding is involved in the pathogenesis of endometriosis (Fig. 2) (24–30).

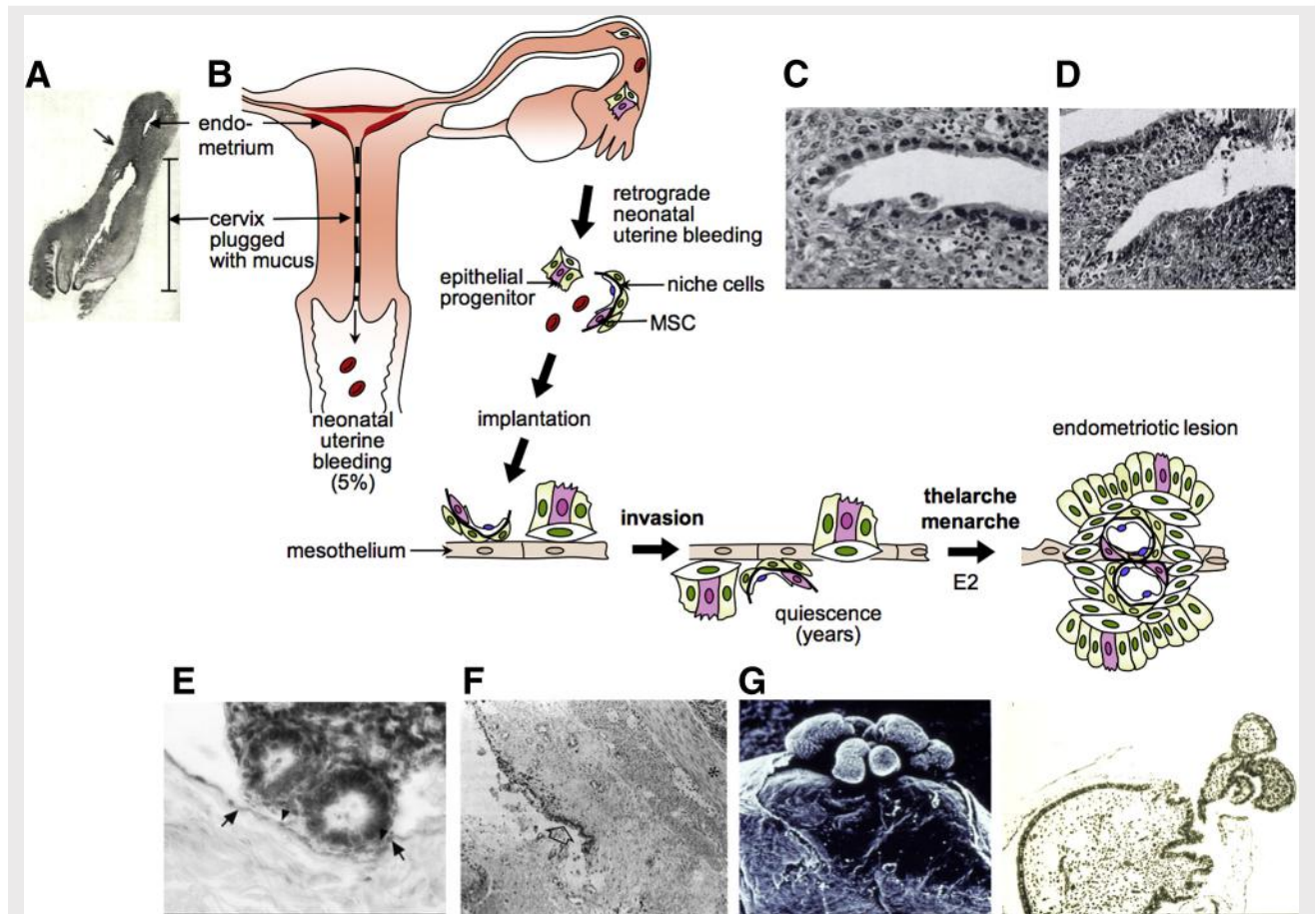
There are three major observations that support the hypothesis of neonatal menstruation and the risk of early

onset endometriosis. Arcellana et al. (31) observed in one neonate with the McKusick-Kaufman syndrome not only the presence of tubal regurgitation of menstrual debris, as also observed by Sampson (6, 7), but in addition, for the first time, the serosal implantation of endometrial fragments. It is important to note that although Sampson observed tubal reflux of menstrual shedding (7) at the time of menstruation, but never reported the early stage of endometrial attachment to the mesothelium and invasion as documented by Witz et al. (32) in experimental conditions. Endometriosis, including the rare premenarcheal endometriosis that presents the same phenotype with angiogenic peritoneal implants and the formation of OMA as adolescent endometriosis, is unexplained by Sampson’s hypothesis (33, 34). A recent review study (35) on endometriosis in symptomatic adolescents has shown that early onset disease is frequently severe and often involves extensive adhesions and even OMAs. In a study of 368 patients with histologically proven deep infiltrating endometriosis Borghese et al. (36) observed that with low birth weight (defined as birth weight <2,500 g) had a higher risk of endometriosis, especially deep infiltrating endometriosis, compared with the reference group. It has been shown that preeclampsia and placental insufficiency increase significantly the risk of the neonatal menstruation (37). Fourth, and probably most important, is the occurrence of the uterine immaturity in the young adolescent. The recent observation (38) of “ontogenetic” uterine P resistance refers to the observation that the endometrial stromal compartment is not intrinsically P responsive at birth. Thus, functional transition of the endometrium to a fully P responsive tissue may be present at birth in newborns showing neonatal uterine bleeding but, in most girls, full endometrial P response will be achieved during adolescence. Hence, the pathogenesis of endometriosis may start in newborns presenting menstrual shedding at a much earlier stage than suggested by the theory of Sampson (7).

The Life Cycle of endometriosis

A life cycle approach of endometriosis reveals unexpected aspects of the natural history of the disease throughout a woman’s life (33). In premenarcheal and adolescent

FIGURE 2



Schematic describing the hypothesis that endometrial stem/progenitor cells may play a role in early onset endometriosis with supporting images from published works. (A) Neonatal uterus and vagina showing relatively long cervix in comparison to the uterine body. The arrow indicates the corpus–cervical junction. Mucus has been removed from the cervix. (B) Schematic showing neonatal uterine bleeding (occurs in 5% of neonates) and hypothesized retrograde neonatal bleeding due to cervical obstruction by thick mucus in the long neonatal cervix. The fragments of shed endometrial tissue are postulated to contain an endometrial epithelial progenitor cell (pink) and a perivascular mesenchymal stem/stromal cell (MSC) (pink) together with niche cells. These rapidly adhere to the neonatal mesothelium, invade and/or become contiguous with the mesothelial lining where they remain in a quiescent state for 10 years. Increasing estrogen (E2) levels associated with thelarche and menarche reactivate the stem/progenitor cells to initiate the growth of endometriosis lesions on the surface of or below the peritoneal mesothelium. Neonatal (C) decidualized and (D) shedding endometrium. (E) Endometrial attachment to the mesothelium occurs within 1 hour and (F) implantation by 18 hours, with endometrial cells becoming contiguous with the mesothelium (arrow) before the onset of quiescence. (G) Scanning electron microscopy (left) and histologic section (right) of a peritoneal endometriotic implant showing a polypoid lesion extending through the mesothelium in a young girl after a decade of quiescence. (Images reprinted with permission [23, 31, 231-233].)

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endometriosis, two types can be distinguished: a classic form that can occur before menarche and a congenital form that is caused by uterine anomaly and outflow obstruction. The lesions include superficial angiogenic peritoneal implants and adhesions, and OMA can occasionally occur. It is suggested that premenarcheal and possibly adolescent endometriosis develop by activation of resting stem cells shed at the time of neonatal retrograde uterine bleeding (29). In adult life, endometriosis can be related to uterine preconditioning by cyclic menstruations acting as a priming mechanism (39). The typical lesions are peritoneal, ovarian, and deep or adenomyotic endometriosis. In postmenopause, endometriosis can develop or be reactivated in the presence or absence of exogenous estrogens

(Es) and can spread to various organs and structures causing obstructive lesions (Supplemental Table 1).

A Disease Characterized by Recurrent Bleeding

Menstruation is widely viewed as serving no purpose other than to reinitiate the menstrual cycle in the absence of pregnancy. Yet, it is striking that cyclic endometrial decidualization followed by menstrual shedding is confined to few species where placentation is characterized by deep trophoblast invasion and remodeling of the spiral arteries from their origin in the inner myometrium. Uterine immaturity in adolescence is manifested by a higher risk of preeclampsia. Apparently, maturation of the uterus to respond adequately

to ovarian hormones by full decidualization is achieved by cyclic menstruation (38). Thus, the emergence of cyclic menstruation may serve to protect uterine tissues from the profound hyperinflammation and oxidative stress associated with deep placentation, a process known as preconditioning. Menstrual preconditioning implies that P withdrawal bleedings or menstruations evolved in the human because of the need to initiate decidualization in the absence of pregnancy and protect uterine tissues from the profound hyperinflammation and oxidative stress that are associated with deep placentation (39). Endometriosis can be viewed as a disorder of exaggerated menstrual preconditioning that confers protection against placentation-related disorders such as preeclampsia.

Whether superficial peritoneal endometriosis, OMA, and deep infiltrating endometriosis represent three different pathogenetic entities or are phenotypes of endometrial tissue in different topographic environments remains to be elucidated. Recurrent ectopic bleeding, however, occurs as a common feature in all three entities. Therefore, the hypothesis was formulated that endometriosis, an ovarian hormone-dependent disorder, is physiologic unless recurrent bleeding in the implant causes progressive disease and symptoms (40). Based on randomized trials (41) against placebo, endometriosis appears to be responsible for chronic pelvic pain symptoms in more than half of confirmed cases. A causal association between severe dysmenorrhea and endometriosis is very probable. This association is independent of the macroscopic type of the lesions or their anatomical locations and may be related to recurrent cyclic microbleeding in the implants (42).

Deep infiltrating endometriosis is found not only in the rectovaginal septum, but also in all fibromuscular pelvic structures such as the uterosacral and utero-ovarian ligaments and the muscular wall of pelvic organs. Cornillie et al. (11) showed that infiltration by endometrial glands and stroma occurs into adjacent fibromuscular tissue along the loose connective tissue and that penetration is arrested at the border of the underlying fat tissue. Hyperplasia of the surrounding smooth muscle and fibrous tissue results in nodule formation and, important, deep infiltrating endometriosis is focally associated with microendometriomas of 500–2,000 μm in diameter particularly in the submucosal layer of the vagina, rectum, or bladder. These non-OMAs are also lined, similarly to the OMA, by endometrial surface epithelium with or without stroma or by polypoid endometriotic tissue.

ReTIAR as Pathogenetic Mechanism of Progressive endometriosis

Whatever the location, the ReTIAR in the formation of both OMA and deep infiltrating endometriosis appears to be the mechanism of progressive endometriosis. The key issue of smooth muscle metaplasia in the pathogenesis of progressive severe endometriosis was first investigated by Hughesdon in 1957 (43). He investigated a series of ovaries with endometrioma in situ and documented the presence of progressive smooth muscle metaplasia (SMM) of the invaginated cortical wall to the extent that in advanced cases the

invaginated pseudocyst was mimicking a uterine structure. Fukunaga (44) confirmed the presence of SMM as a focal, microscopic change or an incomplete rim of SMM surrounding the endometriotic cyst in both the endometrial stroma and the endometriotic cyst. The metaplastic phenomenon in the pathogenesis of deep infiltrating endometriosis, whether located in peritoneum, ovary, rectovaginal septum, or uterosacral ligament was at least partially confirmed by immunohistochemical studies (45). Anaf et al. (45) concluded that the definition of distinct endometriotic entities based on the difference in the tissue composition of the lesions (endometriotic nodules vs. adenomyotic nodules) is inconsistent with the very frequent presence of smooth muscle cells in endometriosis irrespective of its localization. In a review of the literature Donnez et al. (46) suggested that metaplastic changes of Müllerian rests into adenomyotic glands involving the rectovaginal septum and the retroperitoneal space are responsible for the striking proliferation of the smooth muscle, creating an adenomyomatous appearance similar to that of adenomyosis in the endometrium. Van Kaam et al. (47) characterized the fibromuscular tissue and showed that the tissue shared characteristics with pathological wound healing, although the process could not be explained by transforming growth factor- β (TGF- β) alone.

Zhang et al. (48) investigated with *in vitro* experimentations the roles of activated platelets in driving epithelial-to-mesenchymal transition and fibroblast-to-myofibroblast transdifferentiation in endometriosis and concluded that endometriotic lesions and their microenvironment contain all the necessary molecular machinery to promote SMM and fibrogenesis. Their results suggested that endometriotic lesions are wounds that undergo repeated injury and healing, highlighting the importance of platelets in the development of endometriosis.

Therefore, due to the commonality shared with endometriosis (i.e., cyclic bleeding), adenomyotic lesions behave just like endometriotic lesions, which are essentially wounds that undergo ReTIAR. Liu et al. (49) recently explored the difference in the progression of epithelial-to-mesenchymal transition, fibroblast-to-myofibroblast transdifferentiation, SMM, and fibrogenesis within the framework of ReTIAR. They showed that these conditions may share the same pathogenesis/pathophysiology. Proteins that are known to be involved in fibrogenesis, such as THY-1 and PPAR- γ , were also aberrantly expressed in these conditions. The many similarities between OMA and deep infiltrating endometriosis (DIE) indicate that the two conditions share the same pathophysiology and very likely the same pathogenesis. The differences may result from the different lesional microenvironments and, in addition, from the difference in the aging between both as the OMA is the typical severe lesion in adolescent whereas deep endometriosis is typical in the older adult.

The conclusion by Tosti et al. (50) that the specific pathogenic features, such as apoptosis, neuroangiogenesis, oxidative stress, and inflammation, may explain the more severe symptomatology of deep endometriosis should include that OMA is a most severe reproductive disease with an early onset in reproductive life and with great delay in diagnosis. Fortunately, with the advancement in imaging techniques, the

OMA is increasingly diagnosed at an earlier stage when the endometrioma may be less fibrotic and more responsive to medical treatment, making an evaluation of medical options critically important. The fact that OMA causes ovarian fibrosis is a medical problem that should arouse widespread concern in clinicians worldwide. At present, reliable, noninvasive diagnostic procedures of an OMA are available and should be used to identify at the earliest stage the presence of this type of severe pathology.

In conclusion, the different phenotypes (neonatal, premenarcheal, adolescent, adult, menopausal) as well as locations at different sites (superficial, peritoneal, deep, nonperitoneal) of endometriotic lesions raised the question whether they have a different origin and therefore should be considered as separate entities. In light of the natural history of endometriosis (i.e., epithelial-to-mesenchymal transition, fibroblast-to-myofibroblast transdifferentiation, SMM, and finally fibrosis), a pathology-based endometriosis classification system is within reach (51). The present confusion by superficial, deep, pseudodeep, free, enclosed, invasive, metaplastic, or progressive phenotypes can be avoided (52). The two dominant phenotypes of ectopic endometrium largely depend on the localization and have a different sex steroid hormone response (53). The first phenotype characterized by poor sex steroid hormone response is along the Müllerian tract including the uterus, rectovaginal septum, and, to some extent, the uterine ligaments, and the second phenotype outside the Müllerian tract particularly the ovaries. The differentiation has also therapeutic implications, but the current long delay of diagnosis implicates increasing SMM and fibrosis and loss of sex steroid hormone response in all locations. The critical factor in the progression of endometriosis is aging, but unfortunately the long delay in diagnosis of pelvic endometriosis prevents management at the early stage. The neonatal origin in the pathogenesis of early onset endometriosis remains hypothetical, but the hypothesis is falsifiable by the establishment of a registration of neonatal menstruation. Recently several investigators (26, 54) have made a call for systematic registration of the neonatal uterine bleeding.

A POSSIBLE EMBRYONIC ORIGIN?

An alternate theory postulates that endometriosis is caused by defects during organogenesis. Aberrant differentiation and migration of Müllerian ducts cause misplaced spreading of endometrial cells during fetal organogenesis called Müllerianosis (55). Metaplasia of Müllerian remnants was suggested as the histopathogenesis of deep endometriosis in 1992 (56). Autopsy of 36 human fetuses of different gestational age (57) showed the presence of misplaced endometrium at five ectopic sites in 11%. This misplaced endometrium was located at sites correlating with the common location and the incidence of endometriosis in women. Misplaced endometrial glands and embryonic-like duct remnants were also described in six of seven fetuses, supporting the theory that some subtypes of endometriosis are related to an abnormal embryogenesis (58). In a case-controlled study (36) of 743 patients with low birth weight the risk of

developing deep endometriosis is almost two times more, reflecting the possibility of the influence of placental insufficiency on the embryonic development and also favoring the occurrence of neonatal uterine bleeding. A recent study (59), looking at the anogenital distance, a biomarker of prenatal hormonal environment, suggests that endometriosis and especially deep infiltrating endometriosis might have a prenatal origin. Further studies are necessary to prove the importance of hormonal prenatal environment on the later development of endometriosis. Laganà et al. (60) support the hypothesis that ectopic Müllerian remnants from the endometrium, endocervix, and endosalpinx are items from the genital ridge leaked during organogenesis. Special interest should be given to research focusing on the role of genes with a fundamental role in the development of the urogenital tract.

HYPOTHESIS II: DEEP ENDOMETRIOSIS IS A SPECIFIC TYPE OF ABNORMAL ENDOMETRIUM-LIKE CELL, A BENIGN TUMOR

Endometriosis is an enigmatic disease “defined as glands and stroma outside the uterus” with many clinical manifestations and variable symptoms. Its natural history is unclear. In the absence of an animal model mimicking specific endometrial functions such as placentation, and permitting the induction of the different clinical manifestations of endometriosis, the pathophysiology remains debated with hypotheses, theories, and speculation. Neither implantation, metaplasia, lymphatic, nor hematologic spread can explain all clinical manifestations including extrapelvic localizations (61). The genetic/epigenetic changes are proposed as a unifying theory. Key in this discussion is whether the endometriotic cell is similar or different from the endometrium. Whether normal or abnormal, these cells develop in an environment different from the uterus with its specific relationship to the junctional zone. For the many associated events the question remains whether these are cofactors in the development of endometriosis, or rather consequences of the disease.

Clinical Observations

“Endometrial glands and stroma” outside the uterus are not always pathology. As discussed recently (62), microscopic endometriosis in the peritoneum, in the bowel at a distance from a deep endometriotic nodule, and in lymph nodes is not associated with pain or infertility, and there is no evidence so far for their subsequent development.

Endometriosis is—erroneously (63)—considered a progressive disease. However, progression from subtle to more severe lesions, or from typical to cystic or deep, or from cystic to deep has never been observed and remains speculation. In addition, at the moment of clinical diagnosis most lesions are no longer progressive, although they had obviously progressed before. This is a common observation for typical lesions and consistent with their burnt out aspect by pathology. Many cystic lesions can remain unchanged during longer periods of time as evidenced by ultrasound. Most rectovaginal deep endometriosis lesions, which were

followed clinically without surgery, did not progress (clinical observations as acknowledged) (64). However, when traumatized by clinical examination or by puncture for oocyte pick-up, deep endometriosis lesions seem to be activated and might start growing again (63). Deep endometriosis is also an heterogeneous disease. Most lesions are painful, especially during menstruation, but a minority (around 5%) does not cause pain. Some lesions—estimated at <1%—are fast progressive and they do have a different aspect during surgery. Instead of becoming at rest during a pregnancy, some deep lesions do progress and can cause bowel (65, 66), uterus, or bladder perforations. Clinically, typical cystic and deep endometriosis lesions, thus seem to be three different end points as suggested previously (22, 67).

The epidemiology of endometriosis is unclear because of a variable inclusion of subtle lesions and because of diagnostic uncertainties of hospital-based discharge records (64). Clinical observation by surgeons suggests that the prevalence and severity of deep endometriosis is increasing during the past 20 years (64).

Development of (Deep) Endometriosis: Players and Mechanisms

The growth of endometriosis cells varies with the hormonal and immunologic environment. Estrogen and P concentrations in PF (Supplemental Fig. 1) are much higher than in plasma, especially after ovulation (13). These high P concentrations were speculated to inhibit the development of endometriotic implants. Because concentrations were lower in the luteinized unruptured follicle syndrome, it was even speculated that endometriosis development might be a consequence of infertility (8). For many other factors it remains unclear whether they are the cause or the consequence of endometriosis. Pelvic endometriosis is associated with a low-grade inflammation of the peritoneal cavity with higher concentrations of activated macrophages and thus with higher concentrations of the many different cytokines (68–72) and angiogenic factors (73–91). The many immunologic disturbances were recently reviewed (92–99). Specific attention was given to natural killer cells (95, 100) and recently to the role of platelets in their function (101, 102). The natural killer activity is decreased in PF and in plasma of women with endometriosis (103–105) and after excision of deep endometriosis their activity remains low, whereas elevated CA-125 concentrations return to normal (106). This suggests that the immunologic defect might be preexisting to the development of endometriosis. Interestingly, glycoproteins in concentrations as secreted by endometriosis cells in PF (107) decrease the natural killer activity (108), which can be viewed as an autoprotective mechanism of the endometriotic cell. Retrograde menstruation and bleeding in endometriosis lesions generate an iron overload and oxidative stress in the peritoneal cavity (109) and/or in the endometriosis tissue (110). In addition the extremely sensitive mesothelial cells react to retrograde menstruation by mesothelial cell retraction thus facilitating implantation of endometrial cells (111, 112).

Development of (Deep) Endometriosis: What is the Original Cell?

The endometrium is the candidate because shredded menstrual endometrium is viable, was demonstrated in 1927 (113) and has implantation potential as demonstrated by subcutaneous injection (114) as early as 1958. For development on the chicken allantoic membrane, tissue integrity is important (115). It remains unclear whether endometriosis is composed of functional or basal endometrium. The latter might be suggested by the well-demonstrated P resistance (116–123).

The metaplasia theory (5) was proposed because women without a uterus can develop endometriosis. More recently pluripotent stem cells were found in the endometrium and in the peritoneal cavity. Mesothelial-to-mesenchymal transition in the peritoneal cavity is well known (124, 125) with a specific role of platelets (48). Some of these mesenchymal/mesothelial cells of the peritoneal cavity (48), of the mesothelial repair after surgery (126) and in endometrium and endometriosis (127–131) are directly derived from bone marrow. Endometriosis could develop from stem cells in the endometrium (132, 133) or in the peritoneal cavity (29,134–144), possibly induced by genetic changes (60). Recently a specific cell in the endometrium called pale cell (145, 146), because of their appearance, was speculated to be involved in endometriosis and adenomyosis development.

Cells from neonatal retrograde menstruation (54,147–149) or cells remaining from embryonic development (150, 151) are other candidates. Key is that these are (epi)genetically normal cells, and with the actual knowledge of multipotential stem cells in adult life the concept of embryological remnant has become less important.

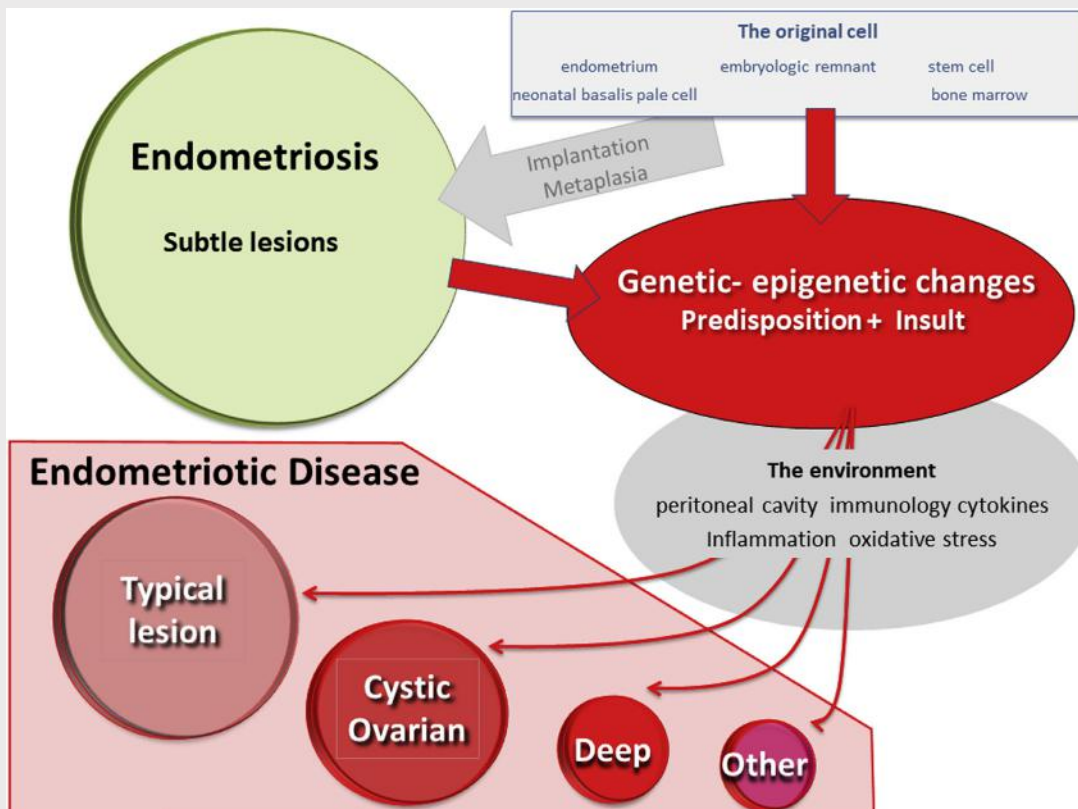
The Endometriotic Disease Theory or Genetic/Epigenetic Changes

The endometriotic disease theory (Fig. 3) was proposed in 1999 (152). Because microscopic and subtle endometriosis lesions are not associated with pain or infertility and most lesions do not progress to more severe pathology, “endometriosis” was suggested for subtle lesions to stress that this was not a clinical pathology. “Endometriotic disease” indicated that typical, cystic, or deep endometriosis was associated with clinical symptoms. To explain the transition from endometriosis to endometriotic disease, the endometriotic disease theory postulated that some cellular or genomic incident or change must have happened. Key in the endometriotic disease theory is that typical, cystic, and deep endometriosis lesions thus should be composed of slightly abnormal cells, similar to many other benign tumors. These cells develop in an abnormal environment outside the uterus and without the specific relationship with the junctional zone. Whether the original cell comes from the endometrium—a specific endometrial cell such as a pale cell, stem cells, bone marrow, or embryonic cells—is not important.

Arguments Pointing to Genetic or Epigenetic Changes

Genetic and epigenetic changes in endometriosis were reviewed recently (20). Endometriosis is an hereditary disease

FIGURE 3



The updated endometriotic disease theory (152). The original cell is not important. Subtle endometriosis by implantation or metaplasia is not a disease until (epi)genetic changes occur. Key is genetic or epigenetic changes, which will lead to typical, cystic, deep, or other lesions as Müllerianosis and extra pelvic localizations. Each of them are (epi)genetically different endometriotic diseases with eventually clinical symptoms.

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and some 50% of endometriosis lesions can be attributed to hereditary factors (153–155). Well established are familial clustering in humans (156) and primates (157). In twin sisters a prevalence of endometriosis (158–161) and age of onset (162) are similar. Prevalence is increased by 6%–9% in first degree relatives (163, 164) and by 15% for severe disease (165, 166).

Genome-wide scanning and linkage analysis have identified potential genes involved. Linkage analysis found two aberrant loci on 10q26 and 7p13–15 (harboring genes such as CYP2C19, INHBA, SFRP4, and HOXA10), but the logarithm of the odds scores are too low to be compatible with one major gene. Genome-wide scanning resulted in 10 significant loci and a significant association of six of these (167, 168). It is too early, however, to understand the exact mechanisms involved. Also attractive is the loss of heterozygosity or the first hit–second hit hypothesis. If in a carrier with a first hit, a second genomic hit would induce endometriosis, this might explain the hereditary character. The many studies that tried to identify a specific hereditary predisposition, especially those investigating detoxication mechanisms, however, failed.

More than 200 differences between the endometrium of women with and without endometriosis were described (for recent reviews, see Refs. [169–173]). These changes might

signal a genetic predisposition. Some differences, however, could be the consequences of endometriosis.

A strong argument is that deep (174, 175) and cystic (176–178) ovarian endometriosis are consistently found to be clonal in origin. This suggests an initial chromosomal or epigenetic change. Typical lesions are too small to be investigated for clonality.

Epigenetic changes, during fetal life (179), have become a focus of interest during the past decade (180–184). They comprise methylation and demethylation of DNA (181, 185, 186), modifications in the histone code in endometriosis tissue in comparison with the endometrium, and experimental modifications of the histone code in cell lines and animal species. Many aberrations have been described, leading to speculation but without a comprehensive view at present.

Dioxin (187–190) and total body radiation (191, 192) might be associated with endometriosis development. Both can have genomic or epigenetic effects (193). In addition the endometriosis developing after total body radiation in primates can have a delay of 5 years, again suggesting a genomic effect.

Taking all elements together we only can speculate why and how some genetic and/or epigenetic changes lead to “abnormal growth of endometrial-like tissue.” External

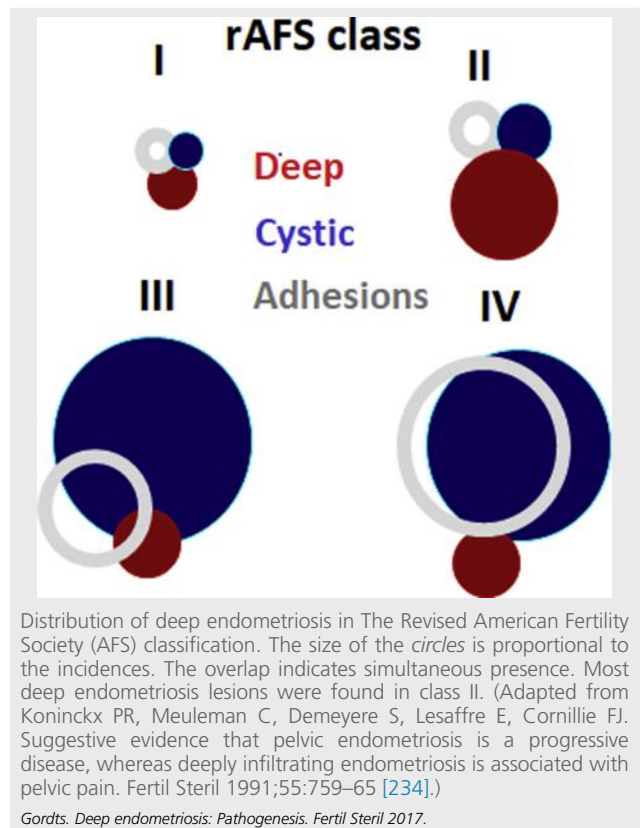
insults and factors, such as oxidative stress in PF (109) or in the tissue because of bleeding, or immunology or cytokines might trigger genetic/epigenetic changes, especially in predisposed cells. This would start the proliferation of endometrium-like tissue into a disease with clinical symptoms. The development into typical, cystic, or deep endometriosis thus will vary with the type of genetic/epigenetic insult. Typical, cystic, and deep endometriosis become three different diseases, which only have endometrium-like tissue in common. Genetic/epigenetic modifications also help to understand the clinical heterogeneity of deep endometriosis.

In conclusion, in the absence of an animal model that permits the induction of severe cystic and deep endometriosis and manipulation of growth, data on pathophysiology are limited to clinical, histologic, and biochemical observations. Whatever the original cell might be, endocrine, immunologic, or biochemical observations cannot explain why some initial lesions will progress and develop into severe endometriosis lesions. The many observations in women with endometriosis can be interpreted as the expression of predisposition and/or as the consequence of endometriosis instead of being the cause. These comprise abundant retrograde menstruation (194) and the recent observations on cellular pathways (195), cytokines (196–199), dendritic cells (200), vitamin D (201), mast cells (202, 203), hypoxia-inducible factor (204), high Mobility Group Box-1 and Toll-Like Receptor 4 (205), matrix metalloproteinase promoter polymorphisms (206), galectin-3 expression (207), promoter polymorphisms of matrix metalloproteinase genes (208), P receptor (PR) expression (209), acylcarnitines, phosphatidylcholines, and sphingomyelins in PF (210), uterine leukocytes (96), vascular epithelial growth factor (211, 212), and other angiogenic factors (213) such as the TGF- β superfamily (214), vezatin expression (215), lymphocytes in blood (216), prostaglandins (217), insulin-like growth factor I (IGF-I) (218), repeated micro-trauma (219) or macro-trauma (63), transcription-3 signaling (220), genetic variants expression (221), and the *Hoxa10/HOXA10* gene (222).

The baboon model with induction of deep endometriosis-like lesions by transplantation of endometrium and myometrium (223) is difficult to interpret. First, spontaneous deep endometriosis has not been reported in primates. Second, it is hardly conceivable that blocks of myometrium and junctional zone/myometrium are the cause of deep endometriosis in humans. These experiments, however, emphasize the importance of tissue integrity—as for the chicken allantoic membrane experiments (115)—and of the special relationship of endometrium and junctional zone for growth and development. Equally intriguing is the role of the increased nerve density and the modulation with time (224, 225). This points to an interaction with the body and can be understood as a cause and as a consequence. It is unclear whether these lesions do undergo (epi)genetic changes as a consequence of their abnormal pelvic environment. Rodent models of endometriosis including human endometrium in nude and SCID mice are not considered appropriate models.

The interpretation that deep endometriosis is a specific disease is reflected in the distribution of deep lesions in all

FIGURE 4



classes of The Revised American Fertility Society (AFS) classification with little association with cystic ovarian endometriosis (Fig. 4). The hypothesis that genetic or epigenetic changes are a prerequisite for development into typical cystic or deep endometriosis is a unifying theory and compatible with all clinical manifestations of endometriosis. In addition it implies that subtle and microscopic lesions are a physiologic condition and that only after genetic/epigenetic changes into typical, cystic, or deep endometriosis does this occur. It is also attractive to consider that the type of genetic/epigenetic changes will orient further development. Deep endometriosis thus becomes one specific type of endometriotic disease.

DISCUSSION

To explain the pathophysiology of endometriosis and more specifically of deep endometriosis, the many hypotheses and observations have been critically reviewed. Two opposing visions persist as evidenced in this review. If the endometriosis cell is considered a normal cell then endometriosis is a single disease and the discussion focuses on the original cell, on subtle lesions as early stages, and on the local and immunologic factors that induce growth, transformation, or metaplasia. If typical, cystic, and deep endometriosis are considered the consequence of a genetic or epigenetic modified cell, microscopic and subtle lesions become a physiologic condition until (epi)genetically modified, the various expressions causing a clinical disease become three or more different

pathologies, and the many associated observations become a consequence of proliferation or a signal of predisposition.

On the other hand, in a life cycle approach of endometriosis it can be suggested that early onset endometriosis develops by activation of resting stem cells shed at the time of neonatal retrograde bleeding to cause angiogenic peritoneal and ovarian endometriotic implants (24, 30). In the adult, the adenomyotic changes, whether in the pelvis or in the ovary, can be related to the uterine preconditioning by cycling menstruation (39). Where cyclic menstruations act as a priming mechanism of uterine preconditioning for deep placentation in case of pregnancy, in case of absence of pregnancy, ReTIAR causing epigenetic modifications may act as a mechanism of deep endometriosis. In this way both hypotheses explain not only the pathogenesis from its earliest stage in the adolescent, but also the progression of endometriosis with aging.

Deep endometriosis, defined as adenomyosis externa, and adenomyosis are morphologically very similar. With Sampson's enunciation of "retrograde menstruation" as a cause of endometriosis the disorder was divorced from adenomyosis and research became focused on how the fragments of menstrual shedding could implant and cause endometriotic pelvic lesions. It is time to reconsider the role of the uterus in the pathogenesis of both disorders as suggested already in 1948 (226)—"one cannot resist the feeling that there is some common denominator between endometrial hyperplasia and adenomyosis, and possibly also pelvic endometriosis." The statement of Novak and De Lima (226) was based on the association of adenomyosis and endometriosis, and observation confirmed more recently by the association of deep endometriosis and adenomyosis. The archimeta concept, the role of the junctional zone (227) with tissue injury and ReTIAR (219, 228) and the role of pale cells could become a unifying concept. The key factor is the local (peritoneal) environment that induces the different types of endometriosis. That E receptors (ERs) and PRs are present not only in glands and stroma but also in the smooth muscle component of deep endometriosis (229), and the expression of α -smooth muscle actin and collagen I in and around endometriotic lesions support the notion of the metaplastic process (230).

The genetic/epigenetic hypothesis is attractive as it is consistent with heredity and clonality and as it explains why not all women with retrograde menstruation and implantation develop endometriosis. The importance of the original cell is weakened by pluripotent stem cells, some of them originating from bone marrow in adult life and by mesothelial-to-mesenchymal transition. The many associated effects are no longer considered the cause but either the consequence of (epi)genetic changes to endometriotic disease or they are factors signaling predisposition. This predisposition can be existing genetic or epigenetic changes facilitating an insult (e.g., differences in the endometrium of women with endometriosis), or biochemical or immunologic factors facilitating growth of endometriotic disease once initiated into typical, cystic, or deep lesions. It is intriguing to realize that, at present, all observations can be explained with this vision. Abundant retrograde menstruation, for example, will

increase peritoneal mesothelial cell retraction and oxidative stress, thus facilitating implantation and subtle lesions and also (epi)genetic changes. Clinically this hypothesis has two consequences. First typical, cystic, and deep endometriosis are three distinct pathologies that only look similar by pathology. Second, although a metaplastic cell will return to normal when the environment changes, (epi)genetically modifications will be transmitted after cell division whatever the environment. Deep endometriosis then becomes a benign tumor.

These complementary views, describing the onset and the further development of (deep) endometriosis lesions are also important to interpret in animal models. They are useful to understand the mechanisms of growth of normal endometrium in an abnormal environment. For the latter reason extrapolation of rodent experiments to the human should be done very carefully. The primate is much closer to the human and thus more appropriate. It cannot be excluded that the induction of deep endometriosis-like lesions by implantation of endometrium and myometrium signals that (epi)genetic modifications did occur (e.g., by the local environment as bleeding with oxidative stress).

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REFERENCES

1. Cullen TS. Adeno-myoma of the round ligament. *Johns Hopkins Hosp Bull* 1896;7:112–4.
2. Cullen TS. Adenoma-myoma uteri diffusum benignum. *Johns Hopkins Hosp Bull* 1896;6:133–7.
3. Cullen TS. The distribution of adenomyomata containing uterine mucosa. *Am J Obstet Gynecol* 1919;80:130–8.
4. Meyer R. Zur frage der Urnieren-genese van Adenomyomen. *Zentralbl Gynakol* 1923;15:577–87.
5. Gruenwald P. Origin of endometriosis from the mesenchyme of the celomic walls. *Am J Obstet Gynecol* 1942;44:470–4.
6. Sampson JA. Heteropic or misplaced endometrial tissue. *Am J Obstet Gynecol* 1925;10:649–64.
7. Sampson JA. Peritoneal endometriosis due to the menstrual dissemination of endometrial tissue into the peritoneal cavity. *Am J Obstet Gynecol* 1927; 14:422–69.
8. Koninckx PR, Ide P, Vandenbroucke W, Brosens IA. New aspects of the pathophysiology of endometriosis and associated infertility. *J Reprod Med* 1980;24:257–60.
9. Halme J, Hammond MG, Hulka JF, Raj SG, Talbert LM. Retrograde menstruation in healthy women and in patients with endometriosis. *Obstet Gynecol* 1984;64:151–4.
10. Koninckx PR. Is mild endometriosis a condition occurring intermittently in all women? *Hum Reprod* 1994;9:2202–5.
11. Cornillie FJ, Oosterlynck D, Lauweryns JM, Koninckx PR. Deeply infiltrating pelvic endometriosis: histology and clinical significance. *Fertil Steril* 1990; 53:978–83.

12. Koninckx PR, Martin DC. Deep endometriosis: a consequence of infiltration or retraction or possibly adenomyosis externa? *Fertil Steril* 1992;58:924–8.
13. Koninckx PR, Heyns W, Verhoeven G, Van Baelen H, Lissens WD, De Moor P, et al. Biochemical characterization of peritoneal fluid in women during the menstrual cycle. *J Clin Endocrinol Metab* 1980;51:1239–44.
14. Anaf V, Simon P, El Nacadi I, Fayt I, Simonart T, Buxant F, et al. Hyperalgesia, nerve infiltration and nerve growth factor expression in deep adenomyotic nodules, peritoneal and ovarian endometriosis. *Hum Reprod* 2002;17:1895–900.
15. Siquara De Sousa AC, Capek S, Amrami KK, Spinner RJ. Neural involvement in endometriosis: Review of anatomic distribution and mechanisms. *Clin Anat* 2015;28:1029–38.
16. Anaf V, El Nakadi N, Simon P, Van de Stadt J, Fayt I, Simonart T, et al. Preferential infiltration of large bowel endometriosis along the nerves of the colon. *Hum Reprod* 2004;19:996–1002.
17. Mechsner S, Weichbrodt M, Riedlinger WF, Bartley J, Kaufmann AM, Schneider A, et al. Estrogen and progesterone receptor positive endometriotic lesions and disseminated cells in pelvic sentinel lymph nodes of patients with deep infiltrating rectovaginal endometriosis: a pilot study. *Hum Reprod* 2008;23:2202–9.
18. Jerman LF, Hey-Cunningham AJ. The role of the lymphatic system in endometriosis: a comprehensive review of the literature. *Biol Reprod* 2015;92:64.
19. Wikipedia contributors. Metaplasia. Available at: <https://en.wikipedia.org/wiki/Metaplasia>. Accessed August 28, 2017.
20. Borghese B, Zondervan KT, Abrao MS, Chapron C, Vaiman D. Recent insights on the genetics and epigenetics of endometriosis. *Clin Genet* 2017;91:254–64.
21. Somigliana E, Infantino M, Candiani M, Vignali M, Chiodini A, Busacca M, et al. Association rate between deep peritoneal endometriosis and other forms of the disease: pathogenetic implications. *Hum Reprod* 2004;19:168–71.
22. Nisolle M, Donnez J. Peritoneal endometriosis, ovarian endometriosis, and adenomyotic nodules of the rectovaginal septum are three different entities. *Fertil Steril* 1997;68:585–96.
23. Ober WB, Bernstein J. Observations on the endometrium and ovary in the newborn. *Pediatrics* 1955;16:445–60.
24. Brosens I, Benagiano G. Is neonatal uterine bleeding involved in the pathogenesis of endometriosis as a source of stem cells? *Fertil Steril* 2013;100:622–3.
25. Bianchi P, Benagiano G, Brosens I. Promoting awareness of neonatal menstruation. *Gynecol Endocrinol* 2017;33:173–8.
26. Puttemans P, Benagiano G, Gargett C, Romero R, Guo SW, Brosens I. Neonatal uterine bleeding as a biomarker for reproductive disorders during adolescence: a worldwide call for systematic registration by nurse midwife. *J Matern Fetal Neonatal Med* 2016;30:1434–6.
27. Brosens I, Gargett CE, Guo SW, Puttemans P, Gordts S, Brosens JJ, et al. Origins and progression of adolescent endometriosis. *Reprod Sci* 2016;23:1282–8.
28. Brosens I, Curcic A, Vejnovic T, Gargett CE, Brosens JJ, Benagiano G. The perinatal origins of major reproductive disorders in the adolescent: Research avenues. *Placenta* 2015;36:341–4.
29. Gargett CE, Schwab KE, Brosens JJ, Puttemans P, Benagiano G, Brosens I. Potential role of endometrial stem/progenitor cells in the pathogenesis of early-onset endometriosis. *Mol Hum Reprod* 2014;20:591–8.
30. Brosens I, Brosens J, Benagiano G. Neonatal uterine bleeding as antecedent of pelvic endometriosis. *Hum Reprod* 2013;28:2893–7.
31. Arcellana RC, Robinson TW, Tyson RW, Joyce MR. McKusick-Kaufman syndrome with legal complications of hydrometrocolpos and congenital endometriosis. *J Perinatol* 1996;16:220–3.
32. Witz CA, Allsup KT, Montoya-Rodriguez IA, Vaughan SL, Centonze VE, Schenken RS. Pathogenesis of endometriosis—current research. *Hum Fertil (Camb)* 2003;6:34–40.
33. Brosens I, Puttemans P, Benagiano G. Endometriosis: a life cycle approach? *Am J Obstet Gynecol* 2013;209:307–16.
34. Marsh EE, Laufer MR. Endometriosis in premenarcheal girls who do not have an associated obstructive anomaly. *Fertil Steril* 2005;83:758–60.
35. Brosens I, Gordts S, Benagiano G. Endometriosis in adolescents is a hidden, progressive and severe disease that deserves attention, not just compassion. *Hum Reprod* 2013;28:2026–31.
36. Borghese B, Sibiude J, Santulli P, Lafay Pillet MC, Marcellin L, Brosens I, et al. Low birth weight is strongly associated with the risk of deep infiltrating endometriosis: results of a 743 case-control study. *PLoS One* 2015;10:e0117387.
37. Levy JM, Rosenthal R, Dellenbach P, Pequenot JP. [Genital crisis in the newborn. repercussion of certain maternal or pregnancy factors on the frequency of neonatal metrorrhagia]. *Arch Franc Ped* 1964;21:819–27.
38. Brosens I, Muter J, Gargett C, Puttemans P, Benagiano G, Brosens JJ. The impact of uterine immaturity on obstetrical syndromes during adolescence. *Am J Obstet Gynecol* 2017;217:546–55.
39. Brosens JJ, Parker MG, McIndoe A, Pijnenborg R, Brosens IA. A role for menstruation in preconditioning the uterus for successful pregnancy. *Am J Obstet Gynecol* 2009;200:615.e1–6.
40. Brosens IA. Endometriosis—a disease because it is characterized by bleeding. *Am J Obstet Gynecol* 1997;176:263–7.
41. Brown J, Farquhar C. Endometriosis: an overview of Cochrane Reviews. *Cochrane Database Syst Rev* 2014;CD009590.
42. Fauconnier A, Chapron C. Endometriosis and pelvic pain: epidemiological evidence of the relationship and implications. *Hum Reprod Update* 2005;11:595–606.
43. Hughesdon PE. The structure of endometrial cysts of the ovary. *J Obstet Gynaecol Br Emp* 1957;64:481–7.
44. Fukunaga M. Smooth muscle metaplasia in ovarian endometriosis. *Histopathology* 2000;36:348–52.
45. Anaf V, Simon P, Fayt I, Noel J. Smooth muscles are frequent components of endometriotic lesions. *Hum Reprod* 2000;15:767–71.
46. Donnez J, van Langendonck A, Casanas-Roux F, van Gossum JP, Pirard C, Jadoul P, et al. Current thinking on the pathogenesis of endometriosis. *Gynecol Obstet Invest* 2002;54(Suppl 1):52–8.
47. Van Kaam KJ, Schouten JP, Nap AW, Dunselman GA, Groothuis PG. Fibromuscular differentiation in deeply infiltrating endometriosis is a reaction of resident fibroblasts to the presence of ectopic endometrium. *Hum Reprod* 2008;23:2692–700.
48. Zhang Q, Duan J, Liu X, Guo SW. Platelets drive smooth muscle metaplasia and fibrogenesis in endometriosis through epithelial-mesenchymal transition and fibroblast-to-myofibroblast transdifferentiation. *Mol Cell Endocrinol* 2016;428:1–16.
49. Liu X, Zhang Q, Guo SW. Histological and immunohistochemical characterization of the similarity and difference between ovarian endometriomas and deep infiltrating endometriosis. *Reprod Sci* 2017 July 18. <http://dx.doi.org/10.1177/19337191171718275>.
50. Tosti C, Pinzauti S, Santulli P, Chapron C, Petraglia F. Pathogenetic mechanisms of deep infiltrating endometriosis. *Reprod Sci* 2015;22:1053–9.
51. Zhang Q, Duan J, Olson M, Fazleabas A, Guo SW. Cellular changes consistent with epithelial-mesenchymal transition and fibroblast-to-myofibroblast transdifferentiation in the progression of experimental endometriosis in baboons. *Reprod Sci* 2016;23:1409–21.
52. Brosens IA. Classification of endometriosis revisited. *Lancet* 1993;341:630.
53. Brosens JJ, Brosens I. Redefining endometriosis: is deep endometriosis a progressive disease? *Hum Reprod* 2000;15:1–3.
54. Gordts S, Puttemans P, Gordts S, Brosens I. Ovarian endometrioma in the adolescent: a plea for early-stage diagnosis and full surgical treatment. *Gynecol Surg* 2015;12:21–30.
55. Batt RE, Smith RA, Buck Louis GM, Martin DC, Chapron C, Koninckx PR, et al. Mullerianosis. *Histol Histopathol* 2007;22:1161–6.
56. Nisolle M, Paindaveine B, Bourdon A, Casanas F, Donnez J. Peritoneal endometriosis: typical aspect and subtle appearance. *Acta Endosc* 1992;22:15–23.
57. Signorile PG, Baldi F, Bussani R, D'Armiento M, de Falco M, Baldi A. Ectopic endometrium in human fetuses is a common event and sustains the theory of mullerianosis in the pathogenesis of endometriosis, a disease that predisposes to cancer. *J Exp Clin Cancer Res* 2009;28:49.

58. Bouquet de JJ, Ayoubi JM, Gianaroli L, Dubuisson JB, Gogusev J, Feki A. Endometriosis: a new cellular and molecular genetic approach for understanding the pathogenesis and evolutivity. *Front Surg* 2014;1:16.
59. Mendiola J, Sanchez-Ferrer ML, Jimenez-Velazquez R, Canovas-Lopez L, Hernandez-Penalver AI, Corbalan-Biyang S, et al. Endometriomas and deep infiltrating endometriosis in adulthood are strongly associated with anogenital distance, a biomarker for prenatal hormonal environment. *Hum Reprod* 2016;31:2377–83.
60. Laganà AS, Vitale SG, Salmeri FM, Triolo O, Frangez HB, Vrtacnik-Bokal E, et al. Unus pro omnibus, omnes pro uno: a novel, evidence-based, unifying theory for the pathogenesis of endometriosis. *Med Hypotheses* 2017;103:10–20.
61. Davis AC, Goldberg JM. Extrapelvic endometriosis. *Semin Reprod Med* 2017;35:98–101.
62. Koninckx PR, Donnez J, Brosens I. Microscopic endometriosis: impact on our understanding of the disease and its surgery. *Fertil Steril* 2016;105:305–6.
63. Canis M, Bourdel N, Houille C, Gremeau AS, Botchorishvili R, Matsuzaki S. Endometriosis may not be a chronic disease: an alternative theory offering more optimistic prospects for our patients. *Fertil Steril* 2016;105:32–4.
64. Koninckx PR, Ussia A, Keckstein J, Wattiez A, Adamyan L. Epidemiology of subtle, typical, cystic, and deep endometriosis: a systematic review. *Gynaecol Surg* 2016;13:457–67.
65. Setubal A, Sidiropoulou Z, Torgal M, Casal E, Lourenco C, Koninckx P. Bowel complications of deep endometriosis during pregnancy or in vitro fertilization. *Fertil Steril* 2014;101:442–6.
66. Vigano P, Corti L, Berlanda N. Beyond infertility: obstetrical and postpartum complications associated with endometriosis and adenomyosis. *Fertil Steril* 2015;104:802–12.
67. Donnez J. Pelvic endometriosis—the same or different entities in disguise? Reply. *Fertil Steril* 1998;70:591–2.
68. Koninckx PR, Kennedy SH, Barlow DH. Endometriotic disease: the role of peritoneal fluid. *Hum Reprod Update* 1998;4:741–51.
69. Barcz E, Kaminski P, Marianowski L. Role of cytokines in pathogenesis of endometriosis. *Med Sci Monit* 2000;6:1042–6.
70. Harada T, Iwabe T, Terakawa N. Role of cytokines in endometriosis. *Fertil Steril* 2001;76:1–10.
71. Wu MY, Ho HN. The role of cytokines in endometriosis. *Am J Reprod Immunol* 2003;49:285–96.
72. Kyama CM, Mihalyi A, Simsa P, Falconer H, Fulop V, Mwenda JM, et al. Role of cytokines in the endometrial-peritoneal cross-talk and development of endometriosis. *Front Biosci (Elite Ed)* 2009;1:444–54.
73. Oosterlynck DJ, Meuleman C, Sobis H, Vandeputte M, Koninckx PR. Angiogenic activity of peritoneal fluid from women with endometriosis. *Fertil Steril* 1993;59:778–82.
74. Morgan KG, Wilkinson N, Buckley CH. Angiogenesis in normal, hyperplastic, and neoplastic endometrium. *J Pathol* 1996;179:317–20.
75. Taylor RN, Ryan IP, Moore ES, Hornung D, Shifren JL, Tseng JF. Angiogenesis and macrophage activation in endometriosis. In: Bullett C, de Ziegler D, Guller S, Levitz M, editors. *Uterus: endometrium and myometrium*, Vol. 828. New York: New York Acad Sciences; 1997: 194–207.
76. Healy DL, Rogers PA, Hii L, Wingfield M. Angiogenesis: a new theory for endometriosis. *Hum Reprod Update* 1998;4:736–40.
77. Matsuzaki S, Canis M, Darcha C, Dechelotte P, Pouly JL, Bruhat MA. Angiogenesis in endometriosis. *Gynecol Obstet Invest* 1998;46:111–5.
78. Fujimoto J, Sakaguchi H, Hirose R, Wen H, Tamaya T. Angiogenesis in endometriosis and angiogenic factors. *Gynecol Obstet Invest* 1999; 48(Suppl 1):14–20.
79. Gazvani R, Templeton A. Peritoneal environment, cytokines and angiogenesis in the pathophysiology of endometriosis. *Reproduction* 2002;123: 217–26.
80. Taylor RN, Lebovic DI, Mueller MD. Angiogenic factors in endometriosis. *Ann N Y Acad Sci* 2002;955:89–100.
81. Becker CM, D'Amato RJ. Angiogenesis and antiangiogenic therapy in endometriosis. *Microvasc Res* 2007;74:121–30.
82. Laschke MW, Menger MD. In vitro and in vivo approaches to study angiogenesis in the pathophysiology and therapy of endometriosis. *Hum Reprod Update* 2007;13:331–42.
83. May K, Becker CM. Endometriosis and angiogenesis. *Minerva Ginecol* 2008;60:245–54.
84. Van LA, Donnez J, Defrere S, Dunselman GA, Groothuis PG. Antiangiogenic and vascular-disrupting agents in endometriosis: pitfalls and promises. *Mol Hum Reprod* 2008;14:259–68.
85. Rogers PA, Donoghue JF, Walter LM, Girling JE. Endometrial angiogenesis, vascular maturation, and lymphangiogenesis. *Reprod Sci* 2009;16: 147–51.
86. Taylor RN, Yu J, Torres PB, Schickedanz AC, Park JK, Mueller MD, et al. Mechanistic and therapeutic implications of angiogenesis in endometriosis. *Reprod Sci* 2009;16:140–6.
87. Laschke MW, Menger MD. Anti-angiogenic treatment strategies for the therapy of endometriosis. *Hum Reprod Update* 2012;18: 682–702.
88. Machado-Linde F, Pelegrin P, Sanchez-Ferrer ML, Leon J, Cascales P, Parrilla JJ. 2-methoxyestradiol in the pathophysiology of endometriosis: focus on angiogenesis and therapeutic potential. *Reprod Sci* 2012;19: 1018–29.
89. Hey-Cunningham AJ, Peters KM, Zevallos HB, Berbic M, Markham R, Fraser IS. Angiogenesis, lymphangiogenesis and neurogenesis in endometriosis. *Front Biosci (Elite Ed)* 2013;5:1033–56.
90. Djokovic D, Calhaz-Jorge C. Angiogenesis as a therapeutic target in endometriosis. *Acta Med Port* 2014;27:489–97.
91. Mari-Alexandre J, Garcia-Oms J, Barcelo-Molina M, Gilibert-Aguilar J, Estelles A, Braza-Boils A, et al. MicroRNAs and angiogenesis in endometriosis. *Thromb Res* 2015;135(Suppl 1):S38–40.
92. Olovsson M. Immunological aspects of endometriosis: an update. *Am J Reprod Immunol* 2011;66(Suppl 1):101–4.
93. Kobayashi H, Higashiura Y, Shigetomi H, Kajihara H. Pathogenesis of endometriosis: the role of initial infection and subsequent sterile inflammation (Review). *Mol Med Rep* 2014;9:9–15.
94. Kralickova M, Veticka V. Immunological aspects of endometriosis: a review. *Ann Transl Med* 2015;3:153.
95. Thiruchelvam U, Wingfield M, O'Farrelly C. Natural killer cells: key players in endometriosis. *Am J Reprod Immunol* 2015;74:291–301.
96. Parkin KL, Fazleabas AT. Uterine leukocyte function and dysfunction: a hypothesis on the impact of endometriosis. *Am J Reprod Immunol* 2016; 75:411–7.
97. De Barros IBL, Malvezzi H, Gueuvoghlian-Silva BY, Piccinato CA, Rizzo LV, Podgaec S. What do we know about regulatory T cells and endometriosis? A systematic review. *J Reprod Immunol* 2017;120: 48–55.
98. Sikora J, Smycz-Kubanska M, Mielczarek-Palacz A, Kondera-Anasz Z. Abnormal peritoneal regulation of chemokine activation-The role of IL-8 in pathogenesis of endometriosis. *Am J Reprod Immunol* 2017 Jan 25. <http://dx.doi.org/10.1111/aji.12622>. Epub ahead of print.
99. Ahn SH, Monsanto SP, Miller C, Singh SS, Thomas R, Tayade C. Pathophysiology and immune dysfunction in endometriosis. *Biomed Res Int* 2015; 2015:79597.
100. Sikora J, Mielczarek-Palacz A, Kondera-Anasz Z. Role of natural killer cell activity in the pathogenesis of endometriosis. *Curr Med Chem* 2011;18: 200–8.
101. Guo SW, Ding D, Liu X. Anti-platelet therapy is efficacious in treating endometriosis induced in mouse. *Reprod Biomed Online* 2016;33:484–99.
102. Du Y, Liu X, Guo SW. Platelets impair natural killer cell reactivity and function in endometriosis through multiple mechanisms. *Hum Reprod* 2017;32:1–17.
103. Oosterlynck DJ, Cornillie FJ, Waer M, Vandeputte M, Koninckx PR. Women with endometriosis show a defect in natural killer activity resulting in a decreased cytotoxicity to autologous endometrium. *Fertil Steril* 1991;56: 45–51.
104. Oosterlynck DJ, Meuleman C, Waer M, Vandeputte M, Koninckx PR. The natural killer activity of peritoneal fluid lymphocytes is decreased in women with endometriosis. *Fertil Steril* 1992;58:290–5.

105. Tanaka E, Sendo F, Kawagoe S, Hiroi M. Decreased natural killer cell activity in women with endometriosis. *Gynecol Obstet Invest* 1992;34:27–30.
106. Oosterlynck DJ, Meuleman C, Waer M, Koninckx PR. CO₂-laser excision of endometriosis does not improve the decreased natural killer activity. *Acta Obstet Gynecol Scand* 1994;73:333–7.
107. Koninckx PR, Riittinen L, Seppala M, Cornillie FJ. CA-125 and placental protein 14 concentrations in plasma and peritoneal fluid of women with deeply infiltrating pelvic endometriosis. *Fertil Steril* 1992;57:523–30.
108. Okamoto N, Uchida A, Takakura K, Kariya Y, Kanzaki H, Riittinen L, et al. Suppression by human placental protein 14 of natural killer cell activity. *Am J Reprod Immunol* 1991;26:137–42.
109. Donnez J, Binda MM, Donnez O, Dolmans MM. Oxidative stress in the pelvic cavity and its role in the pathogenesis of endometriosis. *Fertil Steril* 2016;106:1011–7.
110. Harlev A, Gupta S, Agarwal A. Targeting oxidative stress to treat endometriosis. *Expert Opin Ther Targets* 2015;1–18.
111. Koninckx PR, Gomel V. Introduction: quality of pelvic surgery and postoperative adhesions. *Fertil Steril* 2016;106:991–3.
112. Koninckx PR, Gomel V, Ussia A, Adamyan L. Role of the peritoneal cavity in the prevention of postoperative adhesions, pain, and fatigue. *Fertil Steril* 2016;106:998–1010.
113. Cron RS, Gey G. The viability of cast-off menstrual endometrium. *Am J Obstet Gynecol* 1927;13:645–7.
114. Ridley JH, Edwards IK. Experimental endometriosis in the human. *Am J Obstet Gynecol* 1958;76:783–90.
115. Nap AW, Groothuis PG, Demir AY, Maas JW, Dunselman GA, de Goeij AF, et al. Tissue integrity is essential for ectopic implantation of human endometrium in the chicken chorioallantoic membrane. *Hum Reprod* 2003;18:30–4.
116. Bulun SE, Cheng YH, Yin P, Imir G, Utsunomiya H, Attar E, et al. Progesterone resistance in endometriosis: link to failure to metabolize estradiol. *Mol Cell Endocrinol* 2006;248:94–103.
117. Wang C, Mavrogianis PA, Fazleabas AT. Endometriosis is associated with progesterone resistance in the baboon (*Papio anubis*) oviduct: evidence based on the localization of oviductal glycoprotein 1 (OVGP1). *Biol Reprod* 2009;80:272–8.
118. Bruner-Tran KL, Ding T, Osteen KG. Dioxin and endometrial progesterone resistance. *Semin Reprod Med* 2010;28:59–68.
119. Bulun SE, Cheng YH, Pavone ME, Yin P, Imir G, Utsunomiya H, et al. 17 β -hydroxysteroid dehydrogenase-2 deficiency and progesterone resistance in endometriosis. *Semin Reprod Med* 2010;28:44–50.
120. Al-Sabbagh M, Lam EW, Brosens JJ. Mechanisms of endometrial progesterone resistance. *Mol Cell Endocrinol* 2012;358:208–15.
121. Barragan F, Irwin JC, Balayan S, Erikson DW, Chen JC, Houshdaran S, et al. Human endometrial fibroblasts derived from mesenchymal progenitors inherit progesterone resistance and acquire an inflammatory phenotype in the endometrial niche in endometriosis. *Biol Reprod* 2016;94:118.
122. Joshi NR, Miyadahira EH, Afshar Y, Jeong JW, Young SL, Lessey BA, et al. Progesterone resistance in endometriosis is modulated by the altered expression of microRNA-29c and FKBP4. *J Clin Endocrinol Metab* 2017;102:141–9.
123. Patel BG, Rudnicki M, Yu J, Shu Y, Taylor RN. Progesterone resistance in endometriosis: origins, consequences and interventions. *Acta Obstet Gynecol Scand* 2017;96:623–32.
124. Sandoval P, Jimenez-Heffernan JA, Guerra-Azcona G, Perez-Lozano ML, Rynne-Vidal A, Albar-Vizcaino P, et al. Mesothelial-to-mesenchymal transition in the pathogenesis of post-surgical peritoneal adhesions. *J Pathol* 2016;239:48–59.
125. Cheng Y, Li L, Wang D, Guo Q, He Y, Liang T, Sun L, et al. Characteristics of Human endometrium-derived mesenchymal stem cells and their tropism to endometriosis. *Stem Cells Int* 2017;2017:4794827.
126. Lucas PA. Stem cells for mesothelial repair: an understudied modality. *Int J Artif Organs* 2007;30:550–6.
127. Fernandez Shaw S, Clarke MT, Hicks B, Naish CE, Barlow DH, Starkey PM. Bone marrow-derived cell populations in uterine and ectopic endometrium. *Hum Reprod* 1995;10:2285–9.
128. Du H, Taylor HS. Contribution of bone marrow-derived stem cells to endometrium and endometriosis. *Stem Cells* 2007;25:2082–6.
129. Zhang WB, Cheng MJ, Huang YT, Jiang W, Cong Q, Zheng YF, et al. A study in vitro on differentiation of bone marrow mesenchymal stem cells into endometrial epithelial cells in mice. *Eur J Obstet Gynecol Reprod Biol* 2012;160:185–90.
130. Sakr S, Naqvi H, Komm B, Taylor HS. Endometriosis impairs bone marrow-derived stem cell recruitment to the uterus whereas bazedoxifene treatment leads to endometriosis regression and improved uterine stem cell engraftment. *Endocrinology* 2014;155:1489–97.
131. Moridi I, Mamillapalli R, Cosar E, Ersoy GS, Taylor HS. Bone marrow stem cell chemotactic activity is induced by elevated CXCL12 in endometriosis. *Reprod Sci* 2016;24:526–33.
132. Savilova AM, Farkhat KN, Yushina MN, Rudimova YV, Makiyan ZN, Adamyan LV. Characteristics of multipotent mesenchymal stromal cells isolated from the endometrium and endometriosis lesions of women with malformations of the internal reproductive organs. *Bull Exp Biol Med* 2017;162:539–44.
133. Gargett CE, Masuda H. Adult stem cells in the endometrium. *Mol Hum Reprod* 2010;16:818–34.
134. Sasson IE, Taylor HS. Stem cells and the pathogenesis of endometriosis. *Ann N Y Acad Sci* 2008;1127:106–15.
135. Oliveira FR, Dela CC, del Puerto HL, Vilamil QT, Reis FM, Camargos AF. Stem cells: are they the answer to the puzzling etiology of endometriosis? *Histol Histopathol* 2012;27:23–9.
136. Mirantes C, Espinosa I, Ferrer I, Dolcet X, Prat J, Matias-Guiu X. Epithelial-to-mesenchymal transition and stem cells in endometrial cancer. *Hum Pathol* 2013;44:1973–81.
137. Gurung S, Deane JA, Masuda H, Maruyama T, Gargett CE. Stem cells in endometrial physiology. *Semin Reprod Med* 2015;33:326–32.
138. Hufnagel D, Li F, Cosar E, Krikun G, Taylor HS. The role of stem cells in the etiology and pathophysiology of endometriosis. *Semin Reprod Med* 2015;33:333–40.
139. Ulukus M. Stem cells in endometrium and endometriosis. *Womens Health (Lond Engl)* 2015;11:587–95.
140. Xu Y, Zhu H, Zhao D, Tan J. Endometrial stem cells: clinical application and pathological roles. *Int J Clin Exp Med* 2015;8:22039–44.
141. Koippallil Gopalakrishnan AR, Kishore U, Madan T. Mesenchymal stem cells: a promising tool for targeted gene therapy of endometriosis. *Regen Med* 2017;12:69–76.
142. Pittatore G, Moggio A, Benedetto C, Bussolati B, Revelli A. Endometrial adult/progenitor stem cells: pathogenetic theory and new antiangiogenic approach for endometriosis therapy. *Reprod Sci* 2014;21:296–304.
143. Maruyama T, Yoshimura Y. Stem cell theory for the pathogenesis of endometriosis. *Front Biosci (Elite Ed)* 2012;4:2854–63.
144. Du H, Taylor HS. Stem cells and reproduction. *Curr Opin Obstet Gynecol* 2010;22:235–41.
145. Ibrahim MG, Chiantera V, Frangini S, Younes S, Kohler C, Taube ET, et al. Ultramicro-trauma in the endometrial-myometrial junctional zone and pale cell migration in adenomyosis. *Fertil Steril* 2015;104:1475–83.
146. Tapmeier TT, Becker CM. Is pale the way to go to understand adenomyosis? *Fertil Steril* 2015;104:1378.
147. Brosens I, Benagiano G. The endometrium from the neonate to the adolescent. *J Matern Fetal Neonatal Med* 2016;29:1195–9.
148. Brosens I, Benagiano G, Brosens JJ. The potential perinatal origin of placental disorders in the young primigravida. *Am J Obstet Gynecol* 2015;212:580–5.
149. Brosens I, Benagiano G. Perinatal origin of endometriosis revisited. *Gynecol Endocrinol* 2015;31:419–21.
150. Makiyan Z. Endometriosis origin from primordial germ cells. *Organogenesis* 2017 May 9:1–8. <http://dx.doi.org/10.1080/15476278.2017.1323162>. [Epub ahead of print.]
151. Signorile PG, Baldi A. Endometriosis: new concepts in the pathogenesis. *Int J Biochem Cell Biol* 2010;42:778–80.
152. Koninckx PR, Barlow D, Kennedy S. Implantation versus infiltration: the Sampson versus the endometriotic disease theory. *Gynecol Obstet Invest* 1999;47(Suppl 1):3–9.

153. Sapkota Y, Attia J, Gordon SD, Henders AK, Holliday EG, Rahmioglu N, et al. Genetic burden associated with varying degrees of disease severity in endometriosis. *Mol Hum Reprod* 2015;21:594–602.
154. Saha R, Pettersson HJ, Svedberg P, Olovsson M, Bergqvist A, Marions L, et al. The heritability of endometriosis. *Fertil Steril* 2015;104:947–52.
155. Baranov VS, Ivaschenko TE, Liehr T, Yarmolinskaya MI. Systems genetics view of endometriosis: a common complex disorder. *Eur J Obstet Gynecol Reprod Biol* 2015;185:59–65.
156. Kennedy SH, Mardon H, Barlow DH. Familial endometriosis. *J Assist Reprod Genet* 1995;12:32–4.
157. Hadfield RM, Yudkin PL, Coe CL, Scheffler J, Uno H, Barlow DH, et al. Risk factors for endometriosis in the rhesus monkey (*Macaca mulatta*): a case-control study. *Hum Reprod Update* 1997;3:109–15.
158. Hadfield RM, Mardon HJ, Barlow DH, Kennedy SH. Endometriosis in monozygotic twins. *Fertil Steril* 1997;68:941–2.
159. Moen MH, Magnus P. The familial risk of endometriosis. *Acta Obstet Gynecol Scand* 1993;72:560–4.
160. Moen MH. Endometriosis in monozygotic twins. *Acta Obstet Gynecol Scand* 1994;73:59–62.
161. Treloar SA, O'Connor DT, O'Connor VM, Martin NG. Genetic influences on endometriosis in an Australian twin sample. *Fertil Steril* 1999;71:701–10.
162. Kennedy S, Hadfield R, Mardon H, Barlow D. Age of onset of pain symptoms in non-twin sisters concordant for endometriosis. *Hum Reprod* 1996;11:403–5.
163. Simpson JL, Elias S, Malinak LR, Buttram VCJ. Heritable aspects of endometriosis. I. Genetic studies. *Am J Obstet Gynecol* 1980;137:327–31.
164. Coxhead D, Thomas EJ. Familial inheritance of endometriosis in a British population. A case control study. *J Obstet Gynecol* 1993;13:42–4.
165. Kennedy S. The genetics of endometriosis. *J Reprod Med* 1998;43:263–8.
166. Kennedy S, Hadfield R, Westbrook C, Weeks DE, Barlow D, Golding S. Magnetic resonance imaging to assess familial risk in relatives of women with endometriosis. *Lancet* 1998;352:1440–1.
167. Zondervan KT, Rahmioglu N, Morris AP, Nyholt DR, Montgomery GW, Becker CM, et al. Beyond endometriosis genome-wide association study: from genomics to phenomics to the patient. *Semin Reprod Med* 2016;34:242–54.
168. Rahmioglu N, Macgregor S, Drong AW, Hedman AK, Harris HR, Randall JC, et al. Genome-wide enrichment analysis between endometriosis and obesity-related traits reveals novel susceptibility loci. *Hum Mol Genet* 2015;24:1185–99.
169. Carvalho L, Podgaec S, Bellodi-Privato M, Falcone T, Abrao MS. Role of eutopic endometrium in pelvic endometriosis. *J Minim Invasive Gynecol* 2011;18:419–27.
170. Lessey BA, Lebovic DI, Taylor RN. Eutopic endometrium in women with endometriosis: ground zero for the study of implantation defects. *Semin Reprod Med* 2013;31:109–24.
171. Herndon CN, Aghajanova L, Balayan S, Erikson D, Barragan F, Goldfien G, et al. Global transcriptome abnormalities of the eutopic endometrium from women with adenomyosis. *Reprod Sci* 2016;23:1289–303.
172. Laganà AS, Triolo O, Salmeri FM, Granese R, Palmara VI, Ban FH, et al. Natural Killer T cell subsets in eutopic and ectopic endometrium: a fresh look to a busy corner. *Arch Gynecol Obstet* 2016;293:941–9.
173. da Costa e Silva Rde C, Moura KK, Ribeiro Junior CL, Guillo LA. Estrogen signaling in the proliferative endometrium: implications in endometriosis. *Rev Assoc Med Bras (1992)* 2016;62:72–7.
174. Mayr D, Amann G, Siefert C, Diebold J, Andereg B. Does endometriosis really have premalignant potential? A clonal analysis of laser-microdissected tissue. *FASEB J* 2003;17:693–5.
175. Wu Y, Basir Z, Kajdacsy-Balla A, Strawn E, Macias V, Montgomery K, et al. Resolution of clonal origins for endometriotic lesions using laser capture microdissection and the human androgen receptor (HUMARA) assay. *Fertil Steril* 2003;79(Suppl 1):710–7.
176. Tamura M, Fukaya T, Murakami I, Uehara S, Yajima A. Analysis of clonality in human endometrial cysts based on evaluation of X chromosome inactivation in archival formalin-fixed, paraffin-embedded tissue. *Lab Invest* 1998;78:213–8.
177. Yano T, Jimbo H, Yoshikawa H, Tsutsumi O, Taketani Y. Molecular analysis of clonality in ovarian endometrial cysts. *Gynecol Obstet Invest* 1999;47(Suppl 1):41–5.
178. Jimbo H, Hitomi Y, Yoshikawa H, Yano T, Momoeda M, Sakamoto A, et al. Evidence for monoclonal expansion of epithelial cells in ovarian endometrial cysts. *Am J Pathol* 1997;150:1173–8.
179. Kobayashi H, Iwai K, Niuro E, Morioka S, Yamada Y. Fetal programming theory: implication for the understanding of endometriosis. *Hum Immunol* 2014;75:208–17.
180. Guo SW. Epigenetics of endometriosis. *Mol Hum Reprod* 2009;15:587–607.
181. Houshdaran S, Nezhad CR, Vo KC, Zelenko Z, Irwin JC, Giudice LC. Aberrant Endometrial DNA methylome and associated gene expression in women with endometriosis. *Biol Reprod* 2016;95:93.
182. Zelenko Z, Aghajanova L, Irwin JC, Giudice LC. Nuclear receptor, coregulator signaling, and chromatin remodeling pathways suggest involvement of the epigenome in the steroid hormone response of endometrium and abnormalities in endometriosis. *Reprod Sci* 2012;19:152–62.
183. Colon-Caraballo M, Monteiro JB, Flores I. H3K27me3 is an epigenetic mark of relevance in endometriosis. *Reprod Sci* 2015;22:1134–42.
184. Baumann C, Olson M, Wang K, Fazleabas A, de la Fuente R. Arginine methyltransferases mediate an epigenetic ovarian response to endometriosis. *Reproduction* 2015;150:297–310.
185. Koukoura O, Sifakis S, Spandidos DA. DNA methylation in endometriosis (Review). *Mol Med Rep* 2016;13:2939–48.
186. Zidan HE, Rezk NA, Alnemr AA, Abd El Ghany AM. COX-2 gene promoter DNA methylation status in eutopic and ectopic endometrium of Egyptian women with endometriosis. *J Reprod Immunol* 2015;112:63–7.
187. Koninckx PR. The physiopathology of endometriosis: pollution and dioxin. *Gynecol Obstet Invest* 1999;47(Suppl 1):47–9.
188. Rier S, Foster WG. Environmental dioxins and endometriosis. *Semin Reprod Med* 2003;21:145–54.
189. Guo SW, Simsa P, Kyama CM, Mihalyi A, Fulop V, Othman EE, et al. Reassessing the evidence for the link between dioxin and endometriosis: from molecular biology to clinical epidemiology. *Mol Hum Reprod* 2009;15:609–24.
190. Bruner-Tran KL, Osteen KG. Dioxin-like PCBs and endometriosis. *Syst Biol Reprod Med* 2010;56:132–46.
191. Wood DH, Yochmowitz MG, Salmon YL, et al. Proton irradiation and endometriosis. *Aviat Space Environ Med* 1983;54:718–24.
192. Fanton JW, Golden JG. Radiation-induced endometriosis in *Macaca mulatta*. *Radiat Res* 1991;126:141–6.
193. Sofo V, Gotte M, Laganà AS, Salmeri FM, Triolo O, Sturlese E, et al. Correlation between dioxin and endometriosis: an epigenetic route to unravel the pathogenesis of the disease. *Arch Gynecol Obstet* 2015;292:973–86.
194. Barbieri RL. Stenosis of the external cervical os: an association with endometriosis in women with chronic pelvic pain. *Fertil Steril* 1998;70:571–3.
195. Uimari O, Rahmioglu N, Nyholt DR, Vincent K, Missmer SA, Becker C, et al. Genome-wide genetic analyses highlight mitogen-activated protein kinase (MAPK) signaling in the pathogenesis of endometriosis. *Hum Reprod* 2017;32:780–93.
196. Rakhila H, Al-Akoum M, Bergeron ME, Leboeuf M, Lemyre M, Akoum A, et al. Promotion of angiogenesis and proliferation cytokines patterns in peritoneal fluid from women with endometriosis. *J Reprod Immunol* 2016;116:1–6.
197. Sapkota Y, Low SK, Attia J, Gordon SD, Henders AK, Holliday EG, et al. Association between endometriosis and the interleukin 1A (IL1A) locus. *Hum Reprod* 2015;30:239–48.
198. Malutan AM, Drugan C, Walch K, Drugan T, Ciortea R, Mihu D. The association between interleukin-10 (IL-10) -592C/A, -819T/C, -1082G/A promoter polymorphisms and endometriosis. *Arch Gynecol Obstet* 2017;295:503–10.
199. Ahn SH, Edwards AK, Singh SS, Young SL, Lessey BA, Tayade C. IL-17A contributes to the pathogenesis of endometriosis by triggering proinflammatory cytokines and angiogenic growth factors. *J Immunol* 2015;195:2591–600.
200. Izumi G, Koga K, Takamura M, Makabe T, Nagai M, Urata Y, et al. Mannose receptor is highly expressed by peritoneal dendritic cells in endometriosis. *Fertil Steril* 2017;107:167–73.

201. Ingles SA, Wu L, Liu BT, Chen Y, Wang CY, Templeman C, et al. Differential gene expression by 1,25(OH)₂D₃ in an endometriosis stromal cell line. *J Steroid Biochem Mol Biol* 2017;173:223–7.
202. Binda MM, Donnez J, Dolmans MM. Targeting mast cells: a new way to treat endometriosis. *Expert Opin Ther Targets* 2017;21:67–75.
203. Paula R Jr, Oliani AH, Vaz-Oliani DC, D'Avila SC, Oliani SM, Gil CD. The intricate role of mast cell proteases and the annexin A1-FPR1 system in abdominal wall endometriosis. *J Mol Histol* 2015;46:33–43.
204. Zhan L, Wang W, Zhang Y, Song E, Fan Y, Wei B. Hypoxia-inducible factor-1alpha: a promising therapeutic target in endometriosis. *Biochimie* 2016;123:130–7.
205. Yun BH, Chon SJ, Choi YS, Cho S, Lee BS, Seo SK. Pathophysiology of endometriosis: role of high mobility group box-1 and toll-like receptor 4 developing inflammation in endometrium. *PLoS One* 2016;11:e0148165.
206. Ye H, He Y, Wang J, Song T, Lan Z, Zhao Y, et al. Effect of matrix metalloproteinase promoter polymorphisms on endometriosis and adenomyosis risk: evidence from a meta-analysis. *J Genet* 2016;95:611–9.
207. Yang H, Yin J, Ficarrota K, Hsu SH, Zhang W, Cheng C. Aberrant expression and hormonal regulation of Galectin-3 in endometriosis women with infertility. *J Endocrinol Invest* 2016;39:785–91.
208. Yang H, Liu J, Fan Y, Guo Q, Ge L, Yu N, et al. Associations between various possible promoter polymorphisms of MMPs genes and endometriosis risk: a meta-analysis. *Eur J Obstet Gynecol Reprod Biol* 2016;205:174–88.
209. Wolfler MM, Kuppers M, Rath W, Buck VU, Meinhold-Heerlein I, Classen-Linke I. Altered expression of progesterone receptor isoforms A and B in human eutopic endometrium in endometriosis patients. *Ann Anat* 2016;206:1–6.
210. Vouk K, Ribic-Pucelj M, Adamski J, Rizner TL. Altered levels of acylcarnitines, phosphatidylcholines, and sphingomyelins in peritoneal fluid from ovarian endometriosis patients. *J Steroid Biochem Mol Biol* 2016;159:60–9.
211. Liu XJ, Bai XG, Teng YL, Song L, Lu N, Yang RQ. miRNA-15a-5p regulates VEGFA in endometrial mesenchymal stem cells and contributes to the pathogenesis of endometriosis. *Eur Rev Med Pharmacol Sci* 2016;20:3319–26.
212. Young VJ, Ahmad SF, Brown JK, Duncan WC, Horne AW. Peritoneal VEGF-A expression is regulated by TGF-beta1 through an ID1 pathway in women with endometriosis. *Sci Rep* 2015;5:16859.
213. Gogacz M, Galczynski K, Romanek-Piva K, Winkler I, Rechberger T, Adamiak-Godlewska A. [Concentration of selected angiogenic factors in serum and peritoneal fluid of women with endometriosis]. *Ginekol Pol* 2015;86:188–92.
214. Dela CC, Reis FM. The role of TGFbeta superfamily members in the pathophysiology of endometriosis. *Gynecol Endocrinol* 2015;31:511–5.
215. Holdsworth-Carson SJ, Fung JN, Luong HT, Sapkota Y, Bowdler LM, Wallace L, et al. Endometrial vezatin and its association with endometriosis risk. *Hum Reprod* 2016;31:999–1013.
216. Takamura M, Koga K, Izumi G, Hirata T, Harada M, Hirota Y, et al. Simultaneous detection and evaluation of four subsets of CD4+ T lymphocyte in lesions and peripheral blood in endometriosis. *Am J Reprod Immunol* 2015;74:480–6.
217. Sinreih M, Anko M, Kene NH, Kocbek V, Rizner TL. Expression of AKR1B1, AKR1C3 and other genes of prostaglandin F2alpha biosynthesis and action in ovarian endometriosis tissue and in model cell lines. *Chem Biol Interact* 2015;234:320–31.
218. Mu F, Hankinson SE, Schernhammer E, Pollak MN, Missmer SA. A prospective study of insulin-like growth factor 1, its binding protein 3, and risk of endometriosis. *Am J Epidemiol* 2015;182:148–56.
219. Leyendecker G, Bilgicyildirim A, Inacker M, Stalf T, Huppert P, Mall G, et al. Adenomyosis and endometriosis. Re-visiting their association and further insights into the mechanisms of auto-traumatisation. An MRI study. *Arch Gynecol Obstet* 2015;291:917–32.
220. Kim BG, Yoo JY, Kim TH, Shin JH, Langenheim JF, Ferguson SD, et al. Aberrant activation of signal transducer and activator of transcription-3 (STAT3) signaling in endometriosis. *Hum Reprod* 2015;30:1069–78.
221. Fung JN, Holdsworth-Carson SJ, Sapkota Y, Zhao ZZ, Jones L, Girling JE, et al. Functional evaluation of genetic variants associated with endometriosis near GREB1. *Hum Reprod* 2015;30:1263–75.
222. Zanatta A, Rocha AM, Carvalho FM, Pereira RM, Taylor HS, Motta EL, et al. The role of the Hoxa10/HOXA10 gene in the etiology of endometriosis and its related infertility: a review. *J Assist Reprod Genet* 2010;27:701–10.
223. Donnez O, Van Langendonck A, Van Kerk O, Defrere S, Colette S, Van KO, et al. Induction of endometriotic nodules in an experimental baboon model mimicking human deep nodular lesions. *Fertil Steril* 2013;99:783–9.
224. Orellana R, Garcia-Solares J, Donnez J, van Kerk O, Dolmans MM, Donnez O. Important role of collective cell migration and nerve fiber density in the development of deep nodular endometriosis. *Fertil Steril* 2017;107:987–95.e5.
225. Donnez O, Soares M, Defrere S, Dehoux JP, Van Langendonck A, Donnez J, et al. Nerve fiber density in deep nodular endometriotic lesions induced in a baboon experimental model. *Fertil Steril* 2013;100:1144–50.
226. Novak E, de Lima OA. A correlative study of adenomyosis and pelvic endometriosis, with special reference to the hormonal reaction of ectopic endometrium. *Am J Obstet Gynecol* 1948;56:634–44.
227. Leyendecker G, Kunz G. [Endometriosis and adenomyosis]. *Zentralbl Gynaekol* 2005;127:288–94.
228. Leyendecker G, Wildt L, Mall G. The pathophysiology of endometriosis and adenomyosis: tissue injury and repair. *Arch Gynecol Obstet* 2009;280:529–38.
229. Noel JC, Chapron C, Bucella D, Buxant F, Peny MO, Fayt I, et al. Estrogen and progesterone receptors in smooth muscle component of deep infiltrating endometriosis. *Fertil Steril* 2010;93:1774–7.
230. Ibrahim MG, Sillem M, Plendl J, Chiantera V, Sehouli J, Mechsner S. Myofibroblasts are evidence of chronic tissue microtrauma at the endometrial-myometrial junctional zone in uteri with adenomyosis. *Reprod Sci* 2017;10:1410–8.
231. Fluhmann CF. The developmental anatomy of the cervix uteri. *Obstet Gynecol* 1960;15:62–9.
232. Cornillie FJ, Brosens IA, Vasquez G, Riphagen I. Histologic and ultrastructural changes in human endometriotic implants treated with the antiprogesterone steroid ethynorgestrienone (gestrinone) during 2 months. *Int J Gynecol Pathol* 1986;5:95–109.
233. Witz CA, Thomas MR, Montoya-Rodriguez IA, Nair AS, Centonze VE, Schenken RS. Short-term culture of peritoneum explants confirms attachment of endometrium to intact peritoneal mesothelium. *Fertil Steril* 2001;75:385–90.
234. Koninckx PR, Meuleman C, Demeyere S, Lesaffre E, Cornillie FJ. Suggestive evidence that pelvic endometriosis is a progressive disease, whereas deeply infiltrating endometriosis is associated with pelvic pain. *Fertil Steril* 1991;55:759–65.

SUPPLEMENTAL FIGURE 1

